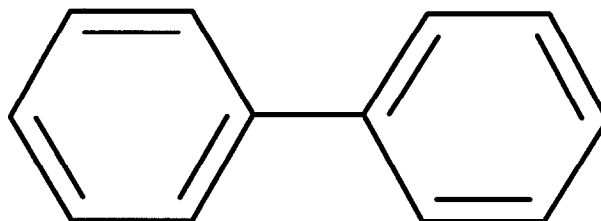


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Biphenyl

CAS Number 92-52-4



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U.S. EPA HPV Challenge Program Submission

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Executive Overview

1,1'-Biphenyl, CAS no. 92-53-4, is an aromatic hydrocarbon that is naturally occurring and is a common combustion product. It is commercially synthesized from benzene or xylene. It is a sublimeable white to yellow crystalline solid with a unique and characteristic odor and a melting point of 70° C. It has low volatility (boiling point 254°C and vapor pressure of 0.0119 hPa @ 25°C) and is relatively insoluble in water (water solubility 7.88 mg/L). Its most extensive use is as a chemical intermediate but it is also used as a heat transfer fluid.

In the environment, based on physicochemical and experimental data, Biphenyl has potential to bioaccumulate (Log K_{ow} = 4.01) and will distribute primarily to soil and water where it will be subject to limited volatilization and rapid biodegradation under conditions favorable to bacteria. It is stable to hydrolysis but expected to react rapidly with atmospheric hydroxyl radicals with a half-life of about 18 hours. Biphenyl is toxic to aquatic species, with an acute LC₅₀ for freshwater fish in the range of 1 to 2 mg/L and daphnia of 0.3 to 1 mg/L; growth inhibition of green alga has also been demonstrated in the range of 1-5 mg/L. The potential for bioaccumulation and adverse effects on aquatic species is offset by its facile biodegradation in the environment.

The acute oral toxicity of Biphenyl is low with an LD₅₀ value of 2400 mg/kg being typically reported for rat gavage studies. Exposure of rats to saturated vapor for 8 hours did not produce any significant adverse effects and the dermal LD₅₀ in rabbits is greater than 2000 mg/kg.

A large number of repeated-dose, subchronic and chronic studies in several species illustrate that Biphenyl is well tolerated orally at lower exposure levels. Repeated dosing, however, at high levels can result in adverse effect to the kidney and bladder with urinary calculus formation. Other than the urinary system, no systemic target organs have been identified. Repeated inhalation of vapor at 50 ppm by rats was found to result in hyperplasia of the trachea; however, exposure to 25 ppm produced only minimal effects.

Adequate *in vitro* tests of genetic toxicity for Biphenyl are available. Multiple *Salmonella typhimurium* reverse mutation assays show lack of mutagenic activity in the presence or absence of metabolic activation and *in vitro* DNA damage studies produce primarily negative results; however, some tests have been positive. The overall preponderance of data suggests that Biphenyl is not genotoxic. .

Developmental toxicity has been investigated using an OECD 414 Guideline-like study in mice and an older, but adequate study in rats. These investigations, both conducted by oral gavage at 0, 100, 250, 500 or 1000 mg/kg-day, indicate that Biphenyl affects the conceptus only at maternally toxic doses, and even at those levels no major malformations occurred. The maternal and developmental NOAEL was found to be 500 mg/kg-day.

The combination of these modern negative developmental toxicity studies with findings from subchronic studies showing lack of effect on reproductive organs fulfills the current requirement for reproductive toxicity information. In addition, there is a three-generation reproduction study and a limited one-generation reproductive investigation in rats that have been reported showing lack of specific reproductive toxicity.

It is concluded that the available information adequately fills all the data elements of the HPV Program for Biphenyl. Conducting further studies would not add significantly to our understanding of this material's hazards or impact guidance that is provided for responsible manufacturing or use of this compound.

Testing Plan and Rationale

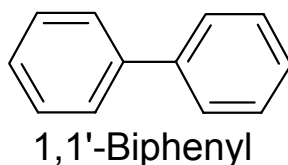
Testing Plan in Tabular Format

CAS Number 92-52-4 1,1'-Biphenyl		Information Available? OECD Study? GLP Study? Supporting Information? Estimation Method? Acceptable? Testing Recommended?						
HPV Endpoint								
Physical Chemical								
Melting Point	Y	N	N	Y	N	Y	N	
Boiling Point	Y	N	N	Y	N	Y	N	
Vapor Pressure	Y	N	N	Y	N	Y	N	
Partition Coefficient	Y	N	N	Y	N	Y	N	
Water Solubility	Y	N	N	Y	N	Y	N	
Environmental & Fate								
Photo-Degradation	Y	N	N	N	Y	Y	N	
Water Stability	Y	N	N	Y	N	Y	N	
Transport	Y	N	N	N	Y	Y	N	
Biodegradation	Y	Y	Y	Y	N	Y	N	
Ecotoxicity								
Acute Fish	Y	N	Y	Y	N	Y	N	
Acute Invertebrate	Y	Y	Y	Y	N	Y	N	
Acute Algae	Y	N	N	Y	N	Y	N	
Toxicity								
Acute	Y	N	N	Y	N	Y	N	
Repeated Dose	Y	N	Y	Y	N	Y	N	
Genetic Toxicology "in vitro"	Y	N	Y	Y	N	Y	N	
Genetic Toxicology "in vivo"	Y	N	Y	Y	N	Y	N	
Reproductive	Y	N	N	Y	N	Y	N	
Developmental	Y	Y	Y	N	N	Y	N	

Introduction

Biphenyl, CAS no 92-52-4, is an aromatic hydrocarbon that is a colorless solid at room temperature and has what is described as a pleasant peculiar odor (1). It is used as an intermediate in the production of a variety of compounds such as: emulsifiers, optical brighteners, crop protection products and plastics, as a dyestuff carrier in textiles and copying paper and as a heat transfer fluid. Biphenyl also occurs naturally in coal tar, crude oil and natural gas (2).

Its structure is shown below:



Biphenyl is also known as (2):

- ☐ Bibenzene
- ☐ 1,1'-Biphenyl
- ☐ Diphenyl
- ☐ 1,1'-Diphenyl
- ☐ Lemonene
- ☐ Phenylbenzene

Exposure in industrial applications is limited by process controls, protective equipment, a very low vapor pressure and excellent warning properties due to its characteristic odor. The ACGIH TLV for Biphenyl is 0.2 ppm.

A broad spectrum of physicochemical, fate and toxicity studies have been conducted on Biphenyl. These studies are briefly reviewed in this testing rationale document, which also describes how these studies meet the SIDS (Screening Information Data Set) end-points of the United States Environmental Protection Agency (USEPA) High Production Volume Challenge (HPV) program. Robust summaries have been prepared for key studies; supporting studies are referenced in these summaries or given as shorter summaries using the IUCLID format. The available data set satisfactorily fulfills the data requirements for the EPA HPV Program. The majority of data elements are filled by high-reliability studies on Biphenyl. Where direct data are not available or data are sparse, surrogates or estimation methods are used to fill the data element. This activity is encouraged by the U.S. EPA and other regulatory authorities to avoid unnecessary testing and animal usage.

Physicochemical Data

Physicochemical data for Biphenyl are available from the literature.

Table 1: Physicochemical Properties of Biphenyl	
Melting Point	69-71° C (1)
Boiling Point	254-255° C @ 1010 hPa (1)
Vapor Pressure	0.0119 hPa @ 25° C (3)
Partition Coefficient	Log $K_{o/w}$ = 4.01 (4)
Water Solubility	7.28 mg/L @ 25° C (5)

These properties indicate that below 70° C, Biphenyl is a volatile solid with low to limited water solubility. The value of the partition coefficient suggests that Biphenyl will partition preferentially into fat; therefore, on the basis of only the octanol-water partition coefficient, Biphenyl is considered to have potential for bioaccumulation; however, if biodegradation and oxidative metabolism are taken into consideration, actual bioaccumulation is much less. The International Program on Chemical Safety (IPCS) has concluded, "...bioaccumulation of the chemical should be of minor importance for aquatic organisms" (6).

Recommendation: No additional physicochemical studies are recommended. The available data fill the HPV required data elements.

Environmental Fate and Pathways

Multiple screening studies using activated sludge as the inoculum have been conducted to assess the biodegradability of Biphenyl. These studies indicate that Biphenyl can be considered readily biodegradable. Biphenyl was tested at 100 mg/L in the MITI test and achieved 66% of the theoretical BOD after two weeks (7). At an initial concentration of 0.8 mg/L, Biphenyl reportedly achieved 100% of the theoretical oxygen uptake in an OECD 301D test (8). In a river die-away study (presented in the robust summaries), Biphenyl at concentrations up to 100 µg/L was shown to undergo almost complete mineralization within a period of eight days without a lag phase (9). Supporting studies showing biodegradability in mixed cultures and with various specific organisms support the ease of Biphenyl's biodegradation (10). It is speculated that the ease with which natural bacteria degrade Biphenyl without a lag period may be related to Biphenyl's natural occurrence in the environment and to its occurrence as a common combustion product.

Biphenyl's photodegradation was estimated using version 1.90 of the Atmospheric Oxidation Program for Microsoft Windows (AOPWIN) that estimates the rate constant for the atmospheric, gas-phase reaction between photochemically produced hydroxyl radicals and organic chemicals. The estimated rate constant is used to calculate atmospheric half-lives for organic compounds based upon average atmospheric concentrations of hydroxyl radical. The program produced an estimated rate constant of $6.8 \text{ E-12 cm}^3/\text{molecule-sec}$; however, the SRC database for hydroxyl radical rate constants (built into AOPWIN) contained an experimental value determined by Atkinson of $7.2 \text{ E-12 cm}^3/\text{molecule-sec}$, which is essentially identical with the calculated value. Using the default atmospheric hydroxyl radical concentration in APOWIN and the experimentally determined rate constant for reaction of Biphenyl with hydroxyl radical, the estimated half-life of Biphenyl vapor in air is approximately 18 hours (see accompanying robust summary for full details).

Water stability has not been quantitatively determined for Biphenyl. Quantitative stability determinations (e.g. OECD 111) are considered unnecessary for compounds containing only non-hydrolysable groups. Under these conditions the SIDS manual states that consideration should be given to using an estimation method. There is no evidence available in the literature that Biphenyl is unstable in water and the structure is that of a simple aromatic hydrocarbon, which is a class of molecule considered to be water unreactive at environmental pH values. The half-life in water is thus estimated as greater than one year. This estimate is confirmed by the review of Harris, who notes specifically that biphenyls as a class are non-hydrolysable (11).

Volatilization and sorption are important in the transport of Biphenyl in aquatic systems. The Henry's Law constant for Biphenyl ($2.5 \times 10^{-4} \text{ atm-m}^3/\text{mol}$) suggests that the molecule may undergo volatilization from aqueous solution. A volatilization half-life of 4.3 hours was estimated for Biphenyl in a stream 1 m deep, flowing 1 m/second, with an air current of 3 meters/second (12).

Theoretical Distribution (Fugacity) of Biphenyl in the environment was estimated using the MacKay EQC level III model with standard defaults in EPIWIN v 3.05. This estimate used the measured vapor pressure of 0.0089 mm Hg, the measured $\log K_{ow}$ of 4.01, an experimentally determined Henry's Law constant, and a measured value for the melting point (13). The results for distribution using a model calculated K_{oc} (adsorption coefficient based on organic carbon content) of 0.0042 and equal initial distribution to air, water and soil are:

○ Air	5.5 %
○ Water	28.8 %
○ Soil	63.8 %
○ Sediment	1.9 %

Recommendation: No additional fate studies are recommended. The available data fill the HPV required elements.

Ecotoxicity

A recent GLP guideline-like study of acute fish toxicity using measured concentrations and flow-through conditions with rainbow trout resulted in an LC₅₀ (192-hour) determination of 1.36 mg/L with a 95% confidence interval of 0.81-1.5 mg/L (14). This finding is in accord with older static tests of Biphenyl on freshwater fish, where 96-h LC₅₀ values from 1.5-4.7 mg/L have been reported, with rainbow trout being the most sensitive species (15). In addition to acute studies, an 87-day early-life-stage study of rainbow trout has been conducted (14). In this study of hatching, development and growth, the NOEC was reported to be 0.229 mg/L Biphenyl and the MATC was assigned as 0.275 mg/L.

Daphnia acute studies run under static conditions have produced a relatively narrow range of toxicity values from an EC₅₀ of 0.73 mg/L in one test (16) to 4.7 mg/L (17) with *Daphnia magna*. The lowest EC₅₀ that has been reported was from a closed flow-through system, where an EC₅₀ of 0.36 mg/L and a NOEC of 0.04 mg/L were reported (18). This acute flow through test was used as a range-finding study for setting Biphenyl concentrations in a reproduction test with *Daphnia magna* in the same closed continuous-flow system. The NOEC after 21 days of incubation including reproductive function was 0.17 mg/L; the maximum-allowable toxicant concentration (MATC) was calculated from this study to be 0.23 mg/L.

An algal growth inhibition study on Biphenyl has been conducted by Hutchinson et al. (19) using two species of green algae *Chlamydomonas angulosa* and *Chlorella vulgaris* that gave 3-hr EC₅₀ values of 1.3 and 3.9 mg/L, respectively. Although a 3-hr EC₅₀ value is a shorter time frame than standard algal growth studies, the abbreviated experimental design of Hutchinson et al. is consistent with the volatilization behavior of Biphenyl in aqueous media. As discussed (*vide supra*), a volatilization half-life of 4.3 hours has been estimated for Biphenyl (20). The 3-hr exposure period with algal combines the exponential growth pattern of algae while maximizing the aqueous exposure of the algae to Biphenyl. As the exposure time is less than one half-life of the estimated aqueous half-life of the chemical (4.3 hours), and as the exposure was conducted using closed flasks, the dose concentrations of Biphenyl in algal media should have fallen by less than 50% from the time zero residues. The measured algal growth 3-hr EC₅₀ value for Biphenyl is consistent with the ECOSAR-predicted value for 96 hours of 1.3 mg/L (see Table 2). This result is also supported by a growth inhibition study of the green alga *Chlorella autotrophica*, which was slightly inhibited (4 mm zone of inhibition) at 1.0 mg Biphenyl/plate and totally inhibited (36 mm zone of inhibition) at 10 mg/plate (20).

Table 2: Comparative Aquatic Toxicity of Biphenyl		
	Reported Values	ECOSAR Prediction
Fish, 96-hour static LC ₅₀ (Rainbow Trout)	1.5 mg/L (15)	1.5 mg/L*
Fish, 192-hour flow through LC ₅₀ (Rainbow Trout)	1.36 mg/L (14)	
Daphnia, 48 hour flow trough EC ₅₀	0.36 mg/L (21)	1.8 mg/L*
Algae, 3-hour EC ₅₀ (<i>Chlamydomonas angulosa</i> and <i>Chlorella vulgaris</i>)	1.3 mg/L (19) 3.9 mg/L	1.3 mg/L*

* Estimated using ECOSAR (22)

Recommendation: The fish, invertebrate, and algal growth inhibition test results are adequate. As ECOSAR-based estimates of toxicity result in an excellent correspondence with the measured values for fish, daphnia, and algae, the overall ecotoxicity data are considered adequate for the purpose of the HPV program.

Metabolism

Facile metabolic conversions of Biphenyl to more polar structures are considered the primary reason why this material does not bioaccumulate to any large extent. The initial metabolites appear to be the same in bacteria as in mammals and these pathways are probably also conserved in fish and invertebrates. In mammals, it has been established that the most prevalent initial metabolite is 4-hydroxybiphenyl. Meyer et al. (23) studied the metabolism of Biphenyl in the rat and reported the primary urinary metabolites as 4-Hydroxybiphenyl (7.7% of dose) and 4,4'-Dihydroxybiphenyl (11.4% of dose). The total urinary recovery 96 hours after administration was 29.5% of the dose and the metabolites detected were conjugates of the mono-, di-, and trihydroxy derivatives of Biphenyl as well as the meta- and para-methyl ethers of the catecholic compounds. These researchers also demonstrated that Biphenyl must be hydroxylated and conjugated prior to biliary excretion and found 5.2% of the dose in the 24 hr bile as conjugates, mainly of 4-hydroxybiphenyl, 4,4'-dihydroxybiphenyl, and 3,4,4'-trihydroxybiphenyl. Other previously undetected minor metabolic products that were identified in the rat were: 3,4'-dihydroxybiphenyl, 3,4,4'-trihydroxybiphenyl, 3,4'-dihydroxy-4-methoxybiphenyl and 4,4'-dihydroxy-3-methoxybiphenyl.

Urinary calculi are a consistent finding in repeated dose studies of Biphenyl at high dose levels, often with males showing a higher incidence than females. A recent study demonstrated that there is a sexual dimorphism in the

composition of the urinary calculi with the male's calculi being composed primarily of potassium 4-hydroxybiphenyl-o-sulfate whereas the calculi in female rats are composed mainly of 4-hydroxybiphenyl and KHSO_4 . Moreover, the calculi have different physical properties and appearance. Photomicrographs and the results of FT-IR analysis indicated that the calculi in males have a multilayer structure consisting of alternating layers of potassium 4-hydroxybiphenyl-o-sulfate and calcium phosphate. In contrast, the calculi in females do not have a multilayer structure, but have open holes in which needle-shaped crystals are sometimes present. This could account for much of the difference in sensitivity between male and female rats.

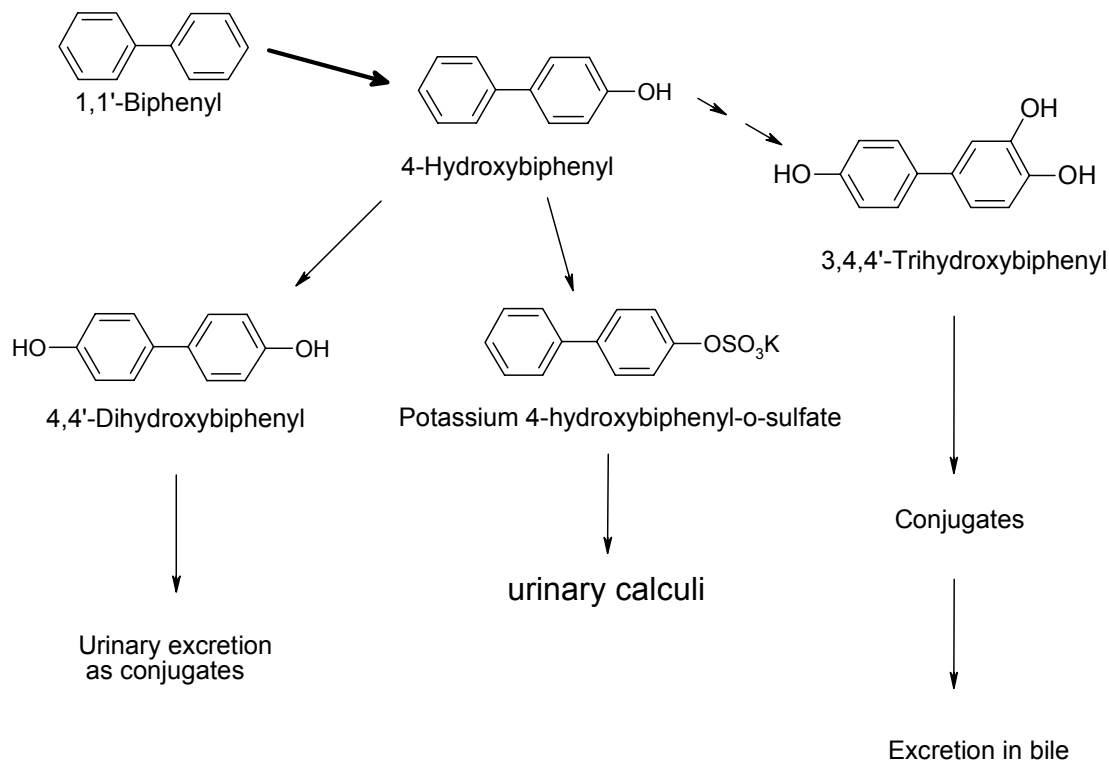


Figure 1: Principle Metabolic Routes of Biphenyl in the Rat

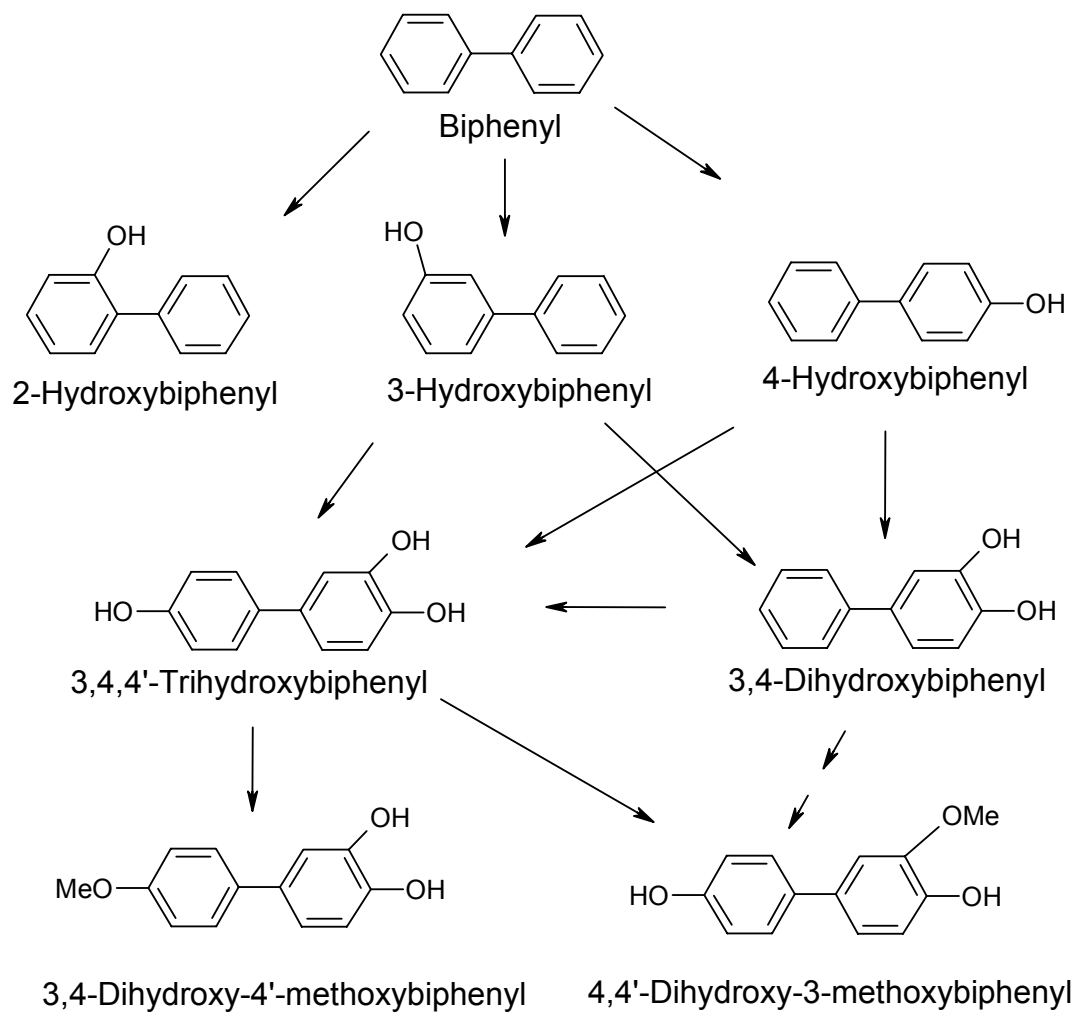


Figure 2: Some of the Known Biphenyl Metabolites

Health Effects

Acute Toxicity

Oral Exposure

Multiple determinations of the oral LD₅₀ of Biphenyl reported with LD₅₀ values ranging from 2180 to 5040 mg/kg indicating a low order of acute oral toxicity for this material. Robust summaries have been prepared from the 1976 and the 1947 studies listed below. The 1949 study gave a lower LD₅₀ but the material was listed as “purity unknown”. A later test by the same laboratory (Mellon Institute) in 1961 using a material described as refined and approximately 99% pure produced a somewhat higher LD₅₀. Overall, the results fall into a reasonably consistent range considering different strains of rats were used with different vehicles and varying purities of test material.

Oral LD ₅₀	Year	Sex studied	Comment	Reference
2180	1949	Male	Purity unknown	24
2400	1976	M & F		25
3280	1947	Not reported		26
3730	1961	Male	Refined material	27
4500	1988			28
5040	1975			29

Table 3. Acute Oral Toxicity of Biphenyl

Inhalation Exposure

No deaths were observed when a group of six female rats were exposed to saturated vapor and mists of purified Biphenyl for 8 hours (27). The actual concentration was not measured but based on the vapor pressure at 20°C and 100° C (5.5 hPa in ECB IUCLID 2000). The vapor concentration is calculated to be in the area of 100 ppm and the aerosol concentration (from condensation of supersaturated vapors) could have been in the range of 20-50 mg/L.

Dermal Exposure

A limited study has indicated that the dermal LD₅₀ of Biphenyl applied to rabbits as a 40% solution/suspension in corn oil, is greater than 5010 mg/kg-bw (25).

Recommendation: No additional acute toxicity studies are recommended. The available data fill the HPV required endpoints for acute toxicity. Although the available studies do not meet all requirements of current OECD guidelines in all cases, the weight of evidence shows the oral and dermal toxicity is very low. Likewise, the limited study of acute saturated vapor by inhalation provides important and scientifically defensible information about vapor toxicity. Conduct of additional studies would not add significantly to our understanding of this material’s toxicity and it is recommended that no additional acute toxicity studies be conducted.

Repeat Dose Toxicity

Multiple repeated dose (14 day through chronic) studies have been conducted with Biphenyl. For the purposes of the HPV program, four have been selected for presentation and summarization. The first is the chronic feeding study by Ambrose (30). The second and third are the 2-year feeding studies in rats and mice conducted by the Japan Bioassay Research Center (31). The final study is a 13-week vapor inhalation study in mice conducted by Cannon Laboratories (32). These were selected because of their duration, relevance of the route of administration, and because they cover two species for carcinogenicity. The Ambrose study is identified as the critical repeated-dose study for the HPV program because of the long duration, the use of several dose levels and the scope of the study (which included two satellite tests of reproductive function).

Oral Exposure

In this chronic feeding study reported by Ambrose et al. (30), 15 rats of each sex were fed diet containing 0, 10, 50, 100, 500, 1000, 5000 or 10000 ppm (0.001 to 1% w/w, ca 0.75, 3.75, 7.5, 37.5, 75, 375 or 750 mg/kg-day). At 5000 ppm, increased liver and kidney weights were observed in females. Concentrations of 5000 and 10000 ppm resulted in shortened lifespan, growth inhibition and lowered hemoglobin values (growth inhibition and reduced hemoglobin levels were attributed to decreased food intake). Treatment related histopathological changes in the kidneys were observed at 5000 ppm and above. The NOAEL was considered 1000 ppm (ca 75 mg/kg-day).

A chronic study using F344/DuCrj rats, performed by the Japan Bioassay Research Center, according to standard protocols, showed a significant increase in neoplastic and non-neoplastic lesions of the urinary bladder and, in high-dose males, a significant increase in calculi within the urinary bladder (31). In this 104-week study, dietary concentrations of Biphenyl were 0, 500, 1500, or 4500 ppm (0, 38, 113, or 338 mg/kg body weight per day). The study report was not available for review; this information was excerpted from the IPCS CICAD document for Biphenyl (6).

In this study, a dose-dependent increase in hyperplasia of the renal pelvis epithelium was reported. Histopathological findings for the kidneys and urinary bladder are summarized in the companion Robust Summary. Other findings included increased serum levels of alkaline phosphatase, aspartate transaminase, and alanine transaminase and an increased urea nitrogen level in low-dose males and mid-dose females, which became more pronounced with increasing doses. Hematological effects were reported in mid- and high-dose females and in high-dose males. A LOEL of 38 mg/kg was derived from these data (it is not clear, however, if this LOEL was assigned by IPCS/WHO or by the original report authors).

A companion chronic study using Crj:BDF1 mice was conducted by the Japan Bioassay Research Center (31). In this study, groups of 50 mice of each sex were given diets containing 0, 667, 2000, or 6000 ppm Biphenyl (0, 100, 300, or 900 mg/kg body weight per day) for 104 weeks prior to sacrifice and complete histopathologic examination. A slight increase in liver tumors (hepatocellular adenomas and carcinomas) and basophilic cell foci of the liver was observed in the females at doses of 300 and 900 mg/kg body weight per day; however, these effects were not concentration dependent and the individual statistical significance was marginal. In male and

female mice, degenerative changes of the nasal cavity respiratory epithelium were reported at doses ≥ 100 mg/kg body weight per day and degenerative changes of the respiratory nasopharynx at doses ≥ 300 mg/kg body weight per day. Other findings included variations in serum enzyme levels (increase in alkaline phosphatase, aspartate transaminase, and alanine transaminase) and an increased urea nitrogen level in the low-dose males and females, which became more pronounced with increasing doses. In female mice receiving ≥ 300 mg Biphenyl/ kg body weight per day and in the high-dose males, degenerative changes in the kidney (increased mineralization of the inner stripe of the outer medulla, increase in desquamation of the epithelium of the renal pelvis) were also observed. High-dose animals also showed reduced body weight gain and food consumption. The study report was not available for review; this information was excerpted from the IPCS CICAD document for Biphenyl (6).

Inhalation Exposure

A 13-week vapor inhalation study using groups of 50 CD-1 mice of each sex exposed to 25 or 50 ppm (160 or 320 mg/m³; analytical concentrations) Biphenyl (32). Exposure was for 7 hours/day, 5 days/week and resulted in hyperaemia and focal hemorrhage in the lung an increase in hyperplasia of the tracheal epithelium. The effects appeared to be dose-related and partially reversible after a 30-day recovery period. In addition, the same laboratory conducted a preliminary 14-day inhalation study under essentially the same conditions and found no effects attributable to the test material (33). Both the 90-day and 14-day studies were limited in scope as only the lungs, trachea, liver, kidneys and spleen were examined microscopically. A robust summary has been prepared for the 90-day study, as it is the only subchronic study available using vapor inhalation as the exposure route. Although the study is limited in scope, it is considered useful in defining the potential of Biphenyl vapor to cause irritation of the respiratory tract.

Recommendation: No additional repeated-dose studies are recommended. The available data fill the HPV required endpoint for repeated-dose toxicity.

Genetic Toxicity

The SIDS/HPV requirement for genetic toxicity screening is for two end-points: generally one test sensitive for point mutation and one sensitive for chromosomal aberrations. In the case of this material, adequate tests have been conducted that cover both of these endpoints.

Genetic Toxicology in vitro

A large number of genotoxicity studies, mostly conducted prior to 1990, have been reported on Biphenyl. The weight of evidence approach suggests that Biphenyl has little genotoxic activity. Results of the in vitro tests are shown in Table 4. Bacterial genotoxicity studies have been uniformly negative while yeast systems have suggested both mutation and mitotic recombination activity. Testing in mammalian cells has produced mixed results with limited positive results for gene mutations and clastogenicity reported only in the presence of metabolic activation.

Test System	End-point	Concentration	Result		References
			N	Y	
<i>Salmonella typhimurium</i>	Reverse mutations	0-5000 µg/plate	-	-	34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48
<i>E.coli</i> WP2, WP2 uvrA-	Gene mutations	0.1-1000 µg/ml	-	-	34, 39, 40
<i>E. coli</i> PQ37	DNA damage	2.4-154 µg/ml	-	-	46
<i>Bacillus subtilis</i> rec assay	DNA damage	no data	-	0	36
<i>Saccharomyces cerevisiae</i> D7	mutat/conversion	≤154 µg/ml	+	+	42
<i>S. cerevisiae</i> D3	Gene conversion	no data	-	-	40, 49
Chinese hamster cells(V79)	Gene mutation	0-100 µg/ml	-	+	48
Mouse lymphoma assay	Gene mutation	0-61 µg/ml	-	(+)	50
Chinese hamster cells (CHL)	Chrom aberration	0-125 µg/ml	-	0	51, 36, 52
Chinese hamster cells(CHL)	Chrom aberration	0-20 µg/ml	-	+	52
Chinese hamster cells (Don)	Chrom aberration	15.4-154 µg/ml	-	0	53
Rat hepatocytes	UDS	0.002-154 µg/ml	0	-	54, 55, 39
Chinese hamster cells(CHL)	SCE	no data	-	0	36
Chinese hamster cells (Don)	SCE	15.4-154 µg/ml	-	0	53
L5178Y cells (DNA unwinding)	DNA damage	0-231 µg/ml	-	+	56
human lung fibroblasts WI-38 cells)	UDS	no data	-	-	40
human fibroblasts ("nick translation assay")	DNA damage	15.4 µg/ml	-	0	57
Y= plus S9, N = no S9, + = positive, (+) = weak positive, - = negative, 0 = no data					

Table 4. In Vitro Genotoxicity Results for Biphenyl

Genetic Toxicology in vivo

Information from genotoxicity studies conducted *in vivo* is limited. In a cytogenetic assay of rat bone marrow cells, the incidence of chromosomal aberrations was reportedly not increased; however, details about the experimental conditions are not available (36). In a second study of bone-marrow chromosome aberrations following inhalation exposure of male rats to an aerosol of 64 or 320 mg Biphenyl/m³ for 30 days (20 exposures), no increase in the frequency of chromosomal aberrations was reported (58). Although the study is lacking certain details, including particle size distribution and cell harvesting times, there is no reason to presume that the results are not valid.

Recommendation: The SIDS requirement for genetic testing has been met as assays sensitive to both point mutation and to clastogenic effects have been conducted using acceptable protocols. No additional genotoxicity testing is recommended.

Reproductive Toxicity

A non-guideline multigenerational study where four successive generations of rats were exposed to dietary levels of 0, 100, 1000 or 10000 ppm Biphenyl has been conducted (59). Although this is an older study the procedure and results are reasonably well documented and it tests the reproductive toxicity of Biphenyl at increasing doses up to those that are clearly maternally and paternally toxic. Marginally reduced fertility occurred at feeding levels that were toxic to the young adult animals as manifest by reduction in weight gains prior to achieving breeding age. Feed levels that were not associated with parental toxicity did not have any effect on reproductive parameters over four generations of exposure. Biphenyl is not considered a specific reproductive toxin to the rat under these conditions. This study was conducted by a scientifically defensible method and its results are congruent with similar dosed feed studies. Because of the duration of the test over three full generations of reproduction, and the marginal effect on measured reproductive parameters, which stayed consistent over the multiple generations, this is considered an adequate test of reproductive toxicity. Additional evidence supporting a lack of reproductive toxicity is found in the 1960 chronic feeding study that incorporated two satellite reproductive and pup survival tests (30).

In addition to the available specific reproductive toxicity data, there are negative developmental toxicity studies (*vide post*). Subchronic studies also found no specific effects on reproductive organs of males or females treated with Biphenyl. For example, as part of the Japan Bioassay Research Center's subchronic study, a detailed gross and microscopic examination of male and female reproductive organs was conducted (31). These studies show that even at systemically toxic doses there is no specific damage to reproductive organs of male or female experimental animals. The available reproductive data and the negative developmental and subchronic studies taken together fulfill the HPV requirement for reproductive toxicity information

Recommendation: No additional reproductive testing is recommended. The available data are sufficient to assess the reproductive toxicity of this material.

Developmental Toxicity

Adequate developmental toxicity studies of Biphenyl have been conducted using both rats (60) and mice (61). The more recent of these studies is an EPA 1984-guideline study using four dose levels and groups of 40 mice per dose level. The results of this investigation conducted by oral gavage at 0, 125, 250, 500 or 1000 mg/kg-day

indicate that Biphenyl is embryotoxic at doses associated with maternal toxicity. The developmental and maternal NOAEL was found to be 500 mg/kg-day with fetotoxicity manifest as early loss. No increase in malformations was observed, even in the presence of maternal toxicity (61). The older study, published in 1979, used groups of 18-20 pregnant Wistar rats dosed by oral gavage at 0, 125, 250, 500 or 1000 mg/kg-day. This study gave a result very similar with the findings in mice; Biphenyl was found to be embryotoxic at doses associated with maternal toxicity. The developmental and maternal NOAEL was found to be 500 mg/kg-day with fetotoxicity manifest as early loss. As was the case with mice, no increase in malformations was observed, even in the presence of maternal toxicity (60). Other supporting information comes from the 1960 chronic-feeding study in rats which had limited reproductive toxicity studies conducted as satellite investigations (30) and from the three-generation study that, although limited in scope, did not indicate any specific developmental toxicity. Thus, there is adequate evidence that Biphenyl is not a specific developmental toxin in rats and mice with dosing conducted by gavage and dosed feed. Taken together, the weight of evidence from these developmental toxicity studies indicates a low developmental toxicity hazard for Biphenyl.

Recommendation: No additional developmental toxicity testing is required as the available data are sufficient to assess the developmental toxicity of this material.

Conclusions

With regard to the parameters specified in the EPA HPV Challenge program, it is concluded that the available information fills all of the requirements for physicochemical parameters, fate information, aquatic toxicity and mammalian toxicity. Although the available studies do not meet all the requirements of the current OECD guidelines in all cases, taken together the information provides a reliable hazard assessment. Conduct of additional studies would not add significantly to our understanding of Biphenyl's toxicity.

References

- 1 O'Neil, MJ (ed.). The Merck Index - An Encyclopedia of Chemicals, Drugs, and Biologicals. Thirteenth edition, Whitehouse Station, NJ: Merck and Co., Inc., 2001
- 2 National Library of Medicine, Hazardous Substance Databank record for Biphenyl CAS Registry Number: 92-52-4, accessed 10/30/2003
- 3 Burkhard, LP et al; J Chem Eng Data 29: 248-50 (1984) as cited in National Library of Medicine Hazardous Substance Data Base, Last Revision Date: 20020806
- 4 Hansch, C., Leo, A., D. Hoekman. Exploring QSAR - Hydrophobic, Electronic, and Steric Constants. Washington, DC: American Chemical Society. 1995. page 97
- 5 Yalkowsky, SH, Dannenfelser, RM; Aquasol Database of Aqueous Solubility. Version 5. College of Pharmacy, University of Arizona-Tucson, AZ. PC Version (1992)
- 6 Concise International Chemical Assessment Document No, 6: Biphenyl. International Program on Chemical Safety, World Health Organization 1999.
- 7 Chemicals Inspection and Testing Institute; Biodegradation and Bioaccumulation Data of Existing Chemicals Based on the CSCL Japan. Japan Chemical Industry Ecology-Toxicology and Information Center. ISBN 4-89074-101-1. as cited in HSDB.
- 8 ECB IUCLID-2000 document for Biphenyl. European Chemicals Bureau, 2000.
- 9 Bailey, RE et al; Biodegradation of the Monochlorophenols and Biphenyl in River Water. Environ Sci Technol 17: 617-21 (1983).
- 10 Freitag, D. Chemosphere 16: 589-98 (1987). Korte F, Klein W; Ecotoxicol Environ Safety 6: 311-27 (1982). Gaffney, PE, J Water Pollut Control Fed 48: 2590-8 (1976). Kitano M, Biodegradation and Bioaccumulation Test on Chemical Substances. OECD Tokyo Meeting. Reference Book 1SU-No. 3 pp. 1-37 (1978). Thom NS, Agg AR; Proc R Soc Lond B189: 347-57 (1975) as cited in HSDB
- 11 Harris, J.C. in Lyman W, Reehl, W and Rosenblat, D.(1990) Handbook of Chemical Property Estimation Methods. American Chemical Society, Washington D.C.
- 12 USEPA. Health and Environmental Effects Profile for 1,1'-biphenyl. Environmental Criteria and Assessment Office, Cincinnati, OH, 35 pp 1984.
- 13 EPIWIN v 3.05, Syracuse Research Corporation, Syracuse NY (April 2000).
- 14 The Dow Chemical Company. Biphenyl: Embryo Larval Toxicity Test With Rainbow Trout, *Salmo Gairdneri* Richardson. Mammalian and Environmental Toxicology Research Laboratory, Final Report. , Study ID: ES-DR-0002-5183-9 02 May 1988.
- 15 BUA Report No. 50, VCH, July 1990. As cited in ECB IUCLID-2000 Sub-ID 92-52-4 in which the following 96-hour static LC50 values were reported: *Lepomis macrochirus*, 4.7 mg/L; *Salmo gairdneri*, 1.5 mg/L.
- 16 Acute Toxicity of Biphenyl to *Daphnia magna*. Report No ES-82-SS-64 Monsanto Environmental Sciences Sept. 3, 1982.

-
- 17 Concise International Chemical Assessment Document No, 6: Biphenyl. International Program on Chemical Safety, World Health Organization 1999. Page 17
 - 18 The Dow Chemical Company, Biphenyl: Flow-Through Chronic Toxicity Test With *Daphnia magna* Straus. Final report. Mammalian and Environmental Toxicology Research Laboratory, Study ID: ES-OR-0002-5183-8, 4 Feb 1988
 - 19 Hutchinson, T.C., J.A. Hellebust, D. Tam, D. Mackay, R.A. Mascarenhas, and W.Y. Shiu. 1980. The correlation of the toxicity to algae of hydrocarbons and halogenated hydrocarbons with their physical-chemical properties. Environ. Sci. Res. 16: 577-586.
 - 20 U.S. Environmental Protection Agency. Health and Environmental Effects Profile for 1,1'-Biphenyl. Environmental Criteria and Assessment Office, Cincinnati, OH 1984.
 - 21 The Dow Chemical Company, Biphenyl: Flow-Through Chronic Toxicity Test With *Daphnia magna* Straus. Mammalian and Environmental Toxicology Research Laboratory, Study ID: ES-OR-0002-5183-8
 - 22 ECOSAR modeling program, version 0.99f, as found in EPIWIN v 3.05, Syracuse Research Corporation, Syracuse NY (April 2000).
 - 23 Meyer T, Scheline RR. The metabolism of Biphenyl. II. Phenolic metabolites in the rat *Acta Pharmacologica et Toxicologica* (1976), 39(4), 419-32
 - 24 Mellon Institute of Industrial Research, Special report on Range Finding Test of Diphenyl, Mellon Institute of Industrial Research Report 12-41 May 5, 1949. From 1983 TSCA 8(d) report of Union Carbide Corp
 - 25 Younger Laboratories Inc. Toxicological Investigations of: Biphenyl. Monsanto Project number Y-76-263. Submitted to Monsanto Co. 8/4/1976
 - 26 Deichmann WB, Kitzmiller KV, Dierker M, and S Witherup. Observations on the Effects of Diphenyl, o- and p-Aminodiphenyl, o- and p-Nitrodiphenyl and Dihydroxyoctachlorodiphenyl Upon Experimental Animals. *J. Ind. Hyg. Toxicol.* 29, 1-13 (1947)
 - 27 Mellon Institute of Industrial Research, Special report on Range Finding Test of Diphenyl, Refined. Mellon Institute of Industrial Research Report 12-41 October 13, 1961. From 1983 TSCA 8(d) report of Union Carbide Corp.
 - 28 Tolstopiatova, G.V. et al.: *Gig. Sanit.* 5: 6-9 (1988) As cited in ECB IUCLID 2000.
 - 29 Prough, R.A. and Burke, M.D.: *Arch. Biochem. Biophys.* 170, 160-168 (1975) As cited in ECB IUCLID 2000
 - 30 Ambrose AM, Booth AN, DeEds F, Cox AJ (1960) A toxicological study of Biphenyl, a citrus fungistat. *Food research*, 25:328-336.
 - 31 Japan Bioassay Research Center (1996) Two year feeding study of Biphenyl in rats and mice. Tokyo, National Institute of Health Sciences (unpublished report). As cited in IPCS CICAD #6 Biphenyl 1999.
 - 32 Cannon Laboratories Inc. 90-day inhalation toxicity study of Biphenyl (99+% purity) in CD mice. sponsored by Sun Co. Inc. November 23, 1977
 - 33 Cannon Laboratories Inc Subacute inhalation toxicity of Biphenyl sponsored by Sun Co. Inc. January 26, 1977
-

-
- 34 Cline JC, McMahon RE (1977) Detection of chemical mutagens. Use of concentration gradient plates in a high capacity screen. *Research communications in chemical pathology and pharmacology*, 16:523-533.
- 35 Purchase IFH, Longstaff E, Ashby J, Styles JA, Anderson D, Lefevre PA, Westwood FR (1978) An evaluation of 6 short-term tests for detecting organic chemical carcinogens. *British journal of cancer*, 37:873-959.
- 36 Kawachi T, Yahagi T, Kada T, Tazima Y, Ishidate M, Sasaki M, Sugiyama T (1980) Cooperative programme on short-term assays for carcinogenicity in Japan. In: Montesano R, Bartsch H, Tomatis L, eds. *Molecular and cellular aspects of carcinogen screening tests*. Lyon, International Agency for Research on Cancer, pp. 323-330 (IARC Scientific Publications No. 27).
- 37 Bronzetti G, Esposito A, Pagano G, Quinto I (1981) A comparative study on the toxicity and mutagenicity of Biphenyl (BP) and diphenyl ether (DPE) in sea urchin, *S. typhimurium* and *S. cerevisiae*. *Mutation research*, 85:233.
- 38 NTP (1980) *Annual plan for fiscal year 1981*. Research Triangle Park, NC, US Department of Health and Human Services, National Toxicology Program, p. 32.
- 39 Probst GS, McMahon RE, Hill LE, Thompson CZ, Epp JK, Neal SB (1981) Chemically-induced unscheduled DNA synthesis in primary rat hepatocyte cultures: a comparison with bacterial mutagenicity using 218 compounds. *Environmental mutagenesis*, 3:11-32.
- 40 Waters MD, Sandhu SS, Simmon VF, Mortelmans KE, Mitchell AD, Jorgenson TA, Jones DCL, Valencia R, Garrett NE (1982) Study of pesticide genotoxicity. *Basic life sciences*, 21:275-326.
- 41 Haworth S, Lawlor T, Mortelmans K, Speck W, Zeiger E (1983) *Salmonella* mutagenicity test results for 250 chemicals. *Environmental mutagenesis*, 5 (Suppl. 1):3-142.
- 42 Pagano G, Esposito A, Giordano GG, Vamvakinos E, Quinto I, Bronzetti G, Bauer C, Corsi C, Nieri R, Ciajolo A (1983) Genotoxicity and teratogenicity of diphenyl and diphenyl ether: a study of sea urchins, yeast, and *Salmonella typhimurium*. *Teratogenesis, carcinogenesis, and mutagenesis*, 3:377-393.
- 43 Pagano G, Cipollaro M, Corsale G, Della Morte R, Esposito A, Giordano GG, Micallo G, Quinto I, Staiano N (1988) Comparative toxicity of diphenyl, diphenyl ester, and some of their hydroxy derivatives. *Médecine Biologie Environnement*, 16:291-297.
- 44 Ishidate M, Sofuni T, Yoshikawa K, Hayashi M, Nohmi T, Sawada M, Matsuoka A (1984) Primary mutagenicity screening of food additives currently used in Japan. *Food and chemical toxicology*, 22:623-636.
- 45 Fujita H, Kojima A, Sasaki M, Hiraga K (1985) Mutagenicity test of antioxidants and fungicides with *Salmonella typhimurium* TA97a, TA102. *Kenkyu Nenpo-Tokyo-toritsu Eisei Kenkyusho*, 36:413-417.
- 46 Brams A, Buchet JP, Crutzen-Fayt MC, de Meester C, Lauwerys R, Leonard A (1987) A comparative study, with 40 chemicals, of the efficiency of the *Salmonella* assay and the SOS chromotest (kit procedure). *Toxicology letters*, 38:123-133.
- 47 Bos RP, Theuws JLG, Jongeneelen FJ, Henderson PT (1988) Mutagenicity of bi-, tri- and tetra-cyclic aromatic hydrocarbons in the "taped-plate assay" and in the conventional *Salmonella* mutagenicity assay. *Mutation research*, 204:203-206.
-

-
- 48 Glatt H, Anklaam E, Robertson LW (1992) Biphenyl and fluorinated derivatives: liver enzyme-mediated mutagenicity detected in *Salmonella typhimurium* and Chinese hamster V79 cells. *Mutation research*, 281:151-156.
- 49 Zimmermann FK, von Borstel RC, von Halle ES, Parry JM, Siebert D, Zetterberg G, Barale R, Loprieno N (1984) Testing of chemicals for genetic activity with *Saccharomyces cerevisiae*: a report of the U.S. Environmental Protection Agency Gene-Tox Program. *Mutation research*, 133:199-244.
- 50 Wangenheim J, Bolcsfoldi G (1988) Mouse lymphoma L5178Y thymidine kinase locus assay of 50 compounds. *Mutagenesis*, 3:193-205.
- 51 Ishidate M, Odashima S (1977) Chromosome tests with 134 compounds on Chinese hamster cells *in vitro* -- a screening for chemical carcinogens. *Mutation research*, 48:337-354.
- 52 Sofuni T, Hayashi M, Matsuoka A, Sawada M, Hatanaka M, Ishidate M (1985) Mutagenicity tests on organic chemical contaminants in city water and related compounds. II. Chromosome aberration tests in cultured mammalian cells. *Eisei Shikensho Hokoku*, 103:64-75.
- 53 Abe S, Sasaki M (1977) Chromosome aberrations and sister chromatid exchanges in Chinese hamster cells exposed to various chemicals. *Journal of the National Cancer Institute*, 58:1635-1641.
- 54 Williams GM (1978) Further improvements in the hepatocyte primary culture DNA repair test for carcinogens: Detection of carcinogenic Biphenyl derivatives. *Cancer letters*, 4:69-75.
- 55 Brouns RE, Poot M, de Vrind R, van Hoek-Kon T, Henderson PT (1979) Measurement of DNA-excision repair in suspensions of freshly isolated rat hepatocytes after exposure to some carcinogenic compounds. Its possible use in carcinogenicity screening. *Mutation research*, 64:425-432.
- 56 Garberg P, Akerblom E-L, Bolcsfoldi G (1988) Evaluation of a genotoxicity test measuring DNA-strand breaks in mouse lymphoma cells by alkaline unwinding and hydroxyapatite elution. *Mutation research*, 203:155-176.
- 57 Snyder RD, Matheson DW (1985) Nick translation -- a new assay for monitoring DNA damage and repair in cultured human fibroblasts. *Environmental mutagenesis*, 7:267-279.
- 58 Dow Chemical Co. (1976) Cytogenetic effects of diphenyl-99 on rat bone marrow cells (EPA Document I.D.: 878213726, received 1983) [cited in BUA, 1994] as cited in IPCS CICAD #6 Biphenyl 1999.
- 59 Stanford Research Institute (undated) Final report - A toxicological study of diphenyl in citrus wraps. Menlo Park, CA EPA Document ID 878213721 OTS # 072253 Received from Dow Chemical Company 06-29-1983
- 60 Khera KS, Whalen C, Angers G, Trivett G (1979) Assessment of the teratogenic potential of piperonyl butoxide, biphenyl, and phosalone in the rat. *Toxicology and applied pharmacology*, 47:353-358.
- 61 Huntingdon Research Centre Ltd., A Study of the Effect of Biphenyl Technical on the Pregnancy of the Mouse. Report THM 1/2/88743, sponsored by Paper Pak Corp, 8/26/1988.
-